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Evaluation of nutmeg (*Myristica fragrans* Houtt) as active component during storage of bovine loins

Evaluación de nuez moscada (*Myristica fragrans* Houtt) como componente activo durante el almacenamiento de lomos de bovino

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ABSTRACT

Meat products are considered highly perishable foods because of their physical, chemical and biological characteristics. Different compounds can be combined to produce active films with greater functionality, which can provide effective protection to food during storage. In this research, active films formulated with polyvinyl alcohol, gelatin, and different oleoresin concentrations of nutmeg (*Myristica fragrans* Houtt) were made by using the casting methodology, a process by which a solution is poured into a mold. Bovine loins (*Longissimus dorsi*) were coated with the active films formulated, stored at 4 °C, performing physicochemical characterization for 1, 3, 6 and 10 days. The results obtained indicate a decrease in pH, humidity percentage, peroxide value (POVs), total volatile nitrogen bases (TVB-N) and greater conservation of the a* parameter (coordinate where +a indicates red, -a indicates green) in loins covered with active films during storage time. Thus, the active films formulated could be used in the meat industry to extend the shelf life of bovine loins.

Keywords: Active films, bases, color, peroxide value, volatile nitrogen.

RESUMEN

Los productos cárnicos son considerados alimentos altamente perecederos debido a sus características físicas, químicas y biológicas. Diferentes compuestos se pueden combinar para la obtención de películas activas con una mayor funcionalidad, que puedan brindar una protección eficaz a los alimentos durante el almacenamiento. En la presente investigación se elaboraron películas activas, formuladas con polivinil alcohol, gelatina y diferentes concentraciones de oleorresina de nuez moscada (*Myristica fragrans* Houtt), empleando la metodología de casting, proceso mediante el cual una solución es vertida en un molde. Lomos de bovino (*Longissimus dorsi*) fueron cubiertos con las películas activas formuladas, almacenados a 4°C, realizando una caracterización fisicoquímica durante 1, 3, 6 y 10 días. Los resultados obtenidos indicaron una disminución en el pH, porcentaje de humedad, índice de peróxido (POVs), bases volátiles nitrogenadas totales (TVB-N) y una mayor conservación del parámetro a* (coordenada donde +a indica rojo, -a indica verde) en los lomos cubiertos con las películas activas, durante el tiempo de almacenamiento. Por lo tanto, las películas activas formuladas podrían ser utilizadas en la industria cárnica para prolongar la vida útil de los lomos de bovino.

Palabras clave: Bases volátiles nitrogenadas, color, índice de peróxidos, películas activas.

INTRODUCTION

Storage time of fresh meat could be extended by using coating agents that delay normal decomposition processes, without modifying the chemical and sensory attributes that characterize it (Antoniewski and Barringer, 2010). Fresh meat and meat conserved under refrigeration conditions (4 - 5°C), during its distribution and consumption, has a limited shelf life between 5 and 6 days due to post mortem autolytic changes from the transformation of the muscle into meat protein and phenomena of lipid oxidation (Zhang et al., 2013). Some studies have demonstrated that shelf life and meat quality can be improved by natural antioxidants added during stages prior to slaughter and post-slaughter (Velasco and Williams, 2011). Antioxidants are compounds that inhibit or delay the oxidation of other molecules or the propagation of oxidizing chain reactions. In meats, lipid oxidation produces rancidity, bad flavor, and unwanted chemical reactions. Recent studies have shown that the active substances contained in nutmeg oleoresins (NO) can act as antioxidants in lipid oxidation in meat (Umaraw and Verna, 2017).

Polymer films can incorporate active substances in their formulation, which turn them into active containers or films for foods. Incorporation of antioxidants and antimicrobials into coating films is carried out to extend the shelf life of foods, delaying lipid oxidation, growth of fungi, yeasts, and bacteria during storage and distribution (Sánchez *et al.*, 2014).

Active films create a gas barrier, reducing permeability and molecule exchange with the environment, thus, avoiding food deterioration; these are characterized by their capacity to act together to improve quality, prolong shelf life, and improve the storage conditions of foods, thus, diminishing the environmental impact produced by plastic materials and increasing sustainability in food industries (Peelman *et al.*, 2013).

Although several conservation methodologies have been studied, currently, prolonging the shelf life of meat continues being a big technological challenge, given the high losses by this costly and nutritional product that reach between 25 and 33% of the total production costs (FAO, 2014) and up to 40% according to that reported by Lorenzo *et al.*, (2014). Hence, the aim of this research was to evaluate the effect of active films with nutmeg on the chemical and microbiological characteristics of bovine loins during storage at 4°C.

MATERIALS AND METHODS

This research was conducted in the Food Laboratory at the Science Interdisciplinary Institute in Universidad del Quindío. To produce the active films, this work followed the methodology described by Figueroa-López et al., (2016a). Initially, an aqueous suspension at 3% p/p of type B gelatin polymer (Bloom 220 - 240) and distilled water at 60°C was homogenized, then the gelatin was added and kept under constant stirring for 30 min at 600rpm. At the same time, an aqueous suspension was prepared with 5% of polyvinyl alcohol (PVA) (Sigma), which was homogenized during 30 min at 90°C. Additionally, a sorbitol solution (99% Sigma-Aldrich) was prepared at 15% heating at 35°C for 15 min. Upon obtaining the solutions, nutmeg oleoresin (NO) (TECNAS, Colombia) was added as active component in concentrations of 0.5, 1.0, and 1.5%, according to the Figueroa et al., (2016b) method. Finally, all the components were mixed for 15 min, distributed over a plastic mold, and dried in an air-circulation stove at 45°C, during 6 h, until obtaining the active films with nutmeg oleoresin (AFNO). As control, a control film (CF) was formulated without adding NO.

To evaluate the effect of the AFNO, these films were used to cover samples of bovine loin cubes (*Longissimus dorsi*) of 2 x 2 x 2cm with a weight of 20g approximately, cut in parallel direction to the muscle fibers, with one day *post mortem* and obtained from the Frigocafé slaughter center in the Armenia city (Quindío), which has INVIMA certification for the slaughter of bovines. As control, a loin was covered with a CF. Finally, the loins were stored at 4°C for their subsequent physical-chemical analysis during 1, 3, 6, and 10 days. The analyses were performed in triplicate and by previously removing the film.

Physical-chemical analysis. The pH measurements were carried out through a Crison PH25[®] portable awl potentiometer (Crison Instruments S.A., Spain) and moisture content $(x^w g/g)$ was determined by desiccation of the samples in a stove at 100°C until the samples reached constant weight (ISO, 1997).

Color analysis. To determine the different color parameters in the bovine loin samples, a Minolta CM-2002R spectrophotometer (Minolta Camera Co., Osaka, Japan) was used, using illuminant D65 and a 10° standard observer (UNE 40-080, 1984). Color determination and expression was conducted based on the CIELAB coordinates and the reflectance values (CIE, 1976). The luminosity (L*), red-green (a*), yellow-blue (b*) parameters were obtained directly from the equipment.

Peroxide value (POVs). Upon finishing the days of storage, the bovine loin samples were analyzed. As target, a sample stored and covered with a CF was used. Lipids were extracted from the loin samples with a mixture of water, methanol, and chloroform (30: 50: 100, v/v). The peroxide values, of the extracts, were measured by the AOAC Method (Horwitz, 2005). The POVs was calculated through the equation (1).

$$POV\left(\frac{meq}{kg}\right) = (S-B) * F * \frac{mol}{L(N)} * \frac{1000}{W}$$

Where *S*, *B*, *F*, mol / L(N), and *W* indicate the volume spent on the titration of the sample, the amount of volume spent on the titration of the target, normality of the Na₂S₂O₃ (0.01 mol equivalent/L), and the sample weight (g), in their order.

Total volatile basic nitrogen (TVB-N). The total content of volatile basic nitrogen in the bovine loin samples stored with AFNO was conducted through distillation, which is the method described by Botta *et al.*, (1984). The amount of TVB-N was calculated through equation (2). The result indicated for each sample was the mean value of both measurements:

$$TVB - N\left(\frac{mg}{100g}\right) = \frac{(v_{1-}v_2) * c * 14}{m * \frac{5}{100}} * 100$$

Where v_1 is the titration volume for the sample analyzed (ml), v_2 is the titration volume in the target (ml), *c* is the concentration of HCl (mol L⁻¹) and *m* is the loin sample weight (g).

Microbiological analysis. The microbiological analysis evaluated loin samples with 1, 3, 6, and 10 days of storage, applying the following methodology: *Staphylococcus aureus* positive coagulase NTC 4779 (ICONTEC, 2007), sulfite-reducing *Clostridium* spores NTC 4834 (ICONTEC, 2000), detection of *Salmonella* NTC 4574 (ICONTEC, 2007), respectively. For Coliform count and detection of *Escherichia coli* was conducted through the procedure described in the NTC 4458 standard (ICONTEC, 2007).

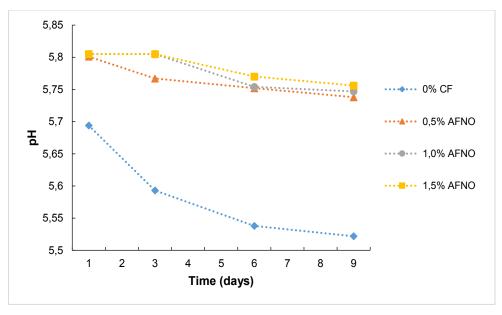
Statistical analysis. A four-factor analysis of variance (ANOVA) was carried out (nutmeg concentration) with four replicates (days of storage) with 95% CI ($p \le 0.05$) where the AFNO and storage time

of the loin samples were the input factors and the response variables were those obtained from the pH, humidity, color (L*, a*, and b*) POVs, and TVB-N analyses. All the experiments were performed in triplicate and were analyzed by using the Stat-graphics Centurion XVII statistical package.

RESULTS AND DISCUSSION

The pH of samples covered with AFNO had a significant decrease (p<0.05) during the days of storage 1, 3, 6, and 10, being more accentuated in the CF, as observed in Figure 1. This behavior may be caused by to dissociations of organic acids, like lactic acid and acetic acid in the muscle tissue of the loins and to the loss of fluid from the meat tissue that causes increased concentration of the solutes and produces decreased pH (Wei *et al.*, 2017).

Significant differences (p < 0.05) were obtained between the final pH of the loin samples and the film used to cover them during storage. This indicates that the AFNO can maintain optimal pH values in the loins during storage.



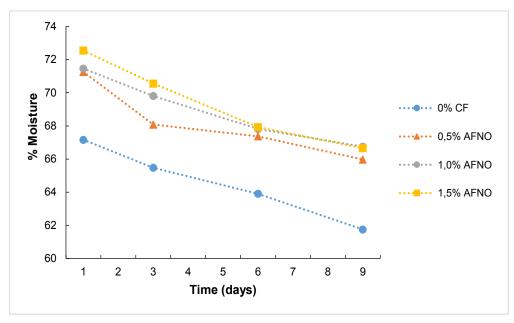
CF = control film; AFNO = Active film with nutmeg oleoresins

Figure 1. pH values against storage time in bovine loins covered with active films with nutmeg oleoresins (AFNO).

These results agree with Šojić *et al.*, (2015), who studied the effect the nutmeg essential oil on the stability of cooked sausage during refrigerated storage, obtaining a decrease in pH during storage; likewise, the sausages had a lower pH value, but significantly higher (p <0.05) compared to sausages used as control.

Loin samples covered with AFNO had a significant decrease in moisture content (p<0.05) with the increase of storage time, being lower than the decrease presented in the CF, as noted in Figure 2. The behavior presented in the loin samples with AFNO was due to the NO which is an oily liquid that acts as a protection barrier, that covers the loin surface, avoids moisture loss during storage, and reduces water migration. Water loss is one of the big problems to processors because it affects the meat weight (Antoniewski and Barringer, 2010). The results agree with Antoniewski et al., (2007), who found that coating of 20% gelatin reduced the purge in beef, pork, and poultry for 14 days under refrigeration.

Color in meat is the principal attribute to determine its freshness. Therefore, it becomes one of the main concerns for processors who seek methods to maintain it. Table 1 shows the results for the CIELAB coordinates of bovine loin samples covered with AFNO during storage at 4°C. Redness (a* value) of the samples with different NO concentrations had significant differences (p <0.05) against samples covered with the control film (CF), while the yellow (b* value) had no significant differences. These results are attributed to the relationship among the color stability, the lipid oxidation and natural pigments (Mancini and Hunt, 2005). Results for luminosity (L*) in the loins covered with the control film presented an increase from 32.703 to 41.093 during storage time, as consequence of the reflection of scattered light due to oxidized lipids (Zaritzky, 2012).



CF = control film; AFNO = Active film with nutmeg oleoresins

Figure 2. Moisture values against storage time in bovine loins covered with active films with nutmeg oleoresins (AFNO).

Sample	L*	a*	b*
One day of storage			
0% NO	35.703 <u>+</u> 0.128	9.828 ± 0.016	4.517 ± 0.013
0.5% AFNO	37.720 ± 0.041	11.787 ± 0.535	6.695 ± 0.069
1.0% AFNO	37.638 ± 0.132	10.576 ± 0.06	5.309 ± 0.035
1.5 AFNO	36.002 ± 0.021	12.776 ± 0.029	7.557 ± 0.039
Three days of storage			
0% NO	33.732 <u>+</u> 0.0052	8.741 ± 0.052	4.658 ± 0.029
0.5% AFNO	36.424 ± 0.024	16.913 ± 0.140	10.11 ± 0.031
1.0% AFNO	36.479 ± 0.092	14.253 ± 0.026	7.594 ± 0.035
1.5 AFNO	35.953 ± 0.004	15.357 ± 0.025	9.372 ± 0.062
Six days of storage			
0% NO	32.305 ± 0.007	9.623 ± 0.093	6.574 ± 0.046
0.5% AFNO	35.450 ± 0.563	16.478 ± 0.259	7.538 ± 0.285
1.0% AFNO	34.755 ± 0.133	12.917 ± 0.054	7.992 ± 0.081
1.5 AFNO	34.435 ± 0.219	15.565 ± 0.054	6.866 ± 0.036
Ten days of storage			
0% NO	31.093 + 0.049	8.909 ± 0.036	6.514 ± 0.058
0.5% AFNO	34.407 ± 0.435	16.413 ± 0.140	6.659 ± 0.149
1.0% AFNO	34.709 ± 0.099	14.192 ± 0.037	9.168 ± 0.021
1.5 AFNO	33.780 ± 0.220	15.483 ± 0.100	8.821 ± 0.084

Table 1. CIELAB coordinates of bovine loin samples

L* = Luminosity, a* = red/green coordinates (+a indicates red, -a indicates green),

b* = yellow/blue coordinates (+b indicates yellow, -b indicates blue)

Changes in luminosity (L*) values obtained upon covering the meat with AFNO over time are shown in Table 1. It, reveals that the L* value in the loins covered with AFNO has no significant changes. The samples covered with CF decreased the value of L* with significant changes (p<0.05) during storage time. This behavior is attributed to the loin samples affected by the loss of moisture, which is translated into lower luminosity. Similar results were reported by Zakaria et al., (2015), who evaluated the effect of the nutmeg extract at different concentrations on the luminosity of raw meat during storage.

The loin samples showed significant differences (p < 0.05), for the a* coordinate, through the nine storage days against loins covered with the CF.

The results obtained agree with Zakaria et al., (2015) who evaluated the effect of the nutmeg extract at different concentrations on the color of raw meat during storage, observing that the different concentrations of nutmeg extract maintained the a* value over time, compared with the control sample. In the same way with Antoniewski et al., (2007) who obtained hight values of a* with increased storage time of the meat, using a gelatin coating. Hence, the AFNO influences on the maintenance of the red tones of the meat as storage time increases. Velasco and Williams (2011) reported that natural antioxidants can delay the loss of meat color extending the red color (a*) and delaying the formation of metamyoglobin that gives meat an unattractive brownish color.

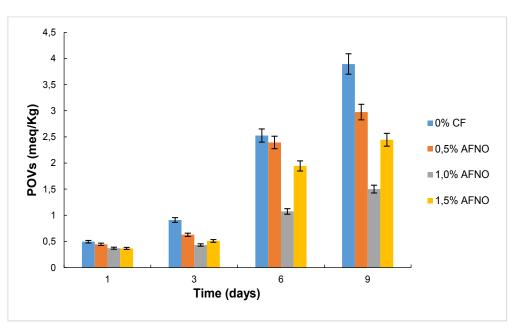
For the b* coordinate no significant differences were observed for the samples covered with AFNO and CF during the storage days . All the samples showed a slight increase in b* values.

The peroxide values reported for all the loin samples had continuous increase during storage time, as noted in Figure 3, and indicate the formation of primary products of lipid oxidation (Yanishlieva and Marinova, 2001).

During storage time, a significant increase (p <0.05) of peroxides was observed in the samples covered with the CF when comparing these with the samples covered with AFNO. This result confirms that the oxygen barrier properties of the films with NO are higher than the films without adding NO during storage time.

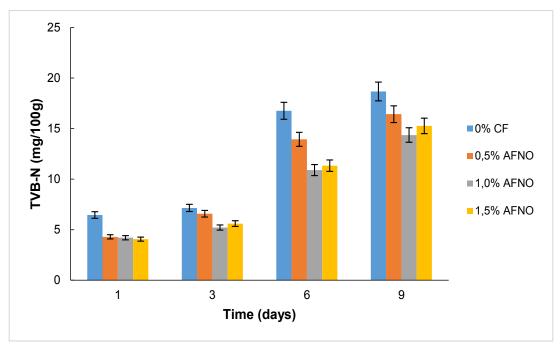
The previous results agree with the studies by Tanabe *et al.*, (2002) who compared antioxidant activities of 22 herbs and culinary species used commonly in lipid oxidation of pork meat. In this case, lipid oxidation was inhibited in the order by nutmeg, caraway, dill, and fennel extracts. The results obtained for the antioxidant activity in the loins were below 10mEq of oxygen/kg of fat), complying with the values established in Resolution 2154/2012.

One of the most-used methodologies to evaluate the degree of meat decomposition is the determination of the TVB-N, which is a key freshness index. Figure 4 displays the results obtained for the TVB-N of the bovine loin samples covered with AFNO and stored at 4°C during 1, 3, 6, and 10 days.



CF = control film; AFNO = Active film with nutmeg oleoresin.

Figure 3. Peroxide values (POVs) in bovine loin samples during storage time.



CF = control film; AFNO = Active film with nutmeg oleoresin

Figure 4. Values of total volatile basic nitrogen (TVB-N) in bovine loin samples during storage time.

The samples covered with AFNO had significant TBV-N differences (p < 0.05) when compared with the samples covered with the CF. The results indicate that the TBV-N values tend to increase with storage time. These were lower than the samples covered with the CF. This behavior indicates the inhibitory effect of the AFNO against secondary oxidation, that is, the delay in releasing nitrogenous substances because of the superior oxygen barrier properties in films with added NO. These results agree with Gómez et al., (2010), who demonstrated that the gelatin and chitosan film with incorporation of nutmeg essential oil was able to reduce the TVB-N value in fish meat during refrigerated storage. The TVB-N values below 20mg N/100g indicate that the loins are acceptable for human consumption (Decree 29210 MAG MEIC; Senasa, 2003).

The microbiological analysis of the loin samples covered with the films obtained counts of

Staphylococcus aureus positive coagulase and sulfite-reducing Clostridium spore count <100 UFC/g and absence of Salmonella during storage time. Likewise, the Escherichia coli count evidenced antimicrobial activity in all the loin samples covered with AFNO and lower total coliform counts than the samples covered with CF. The results obtained are attributed to the antimicrobial activity associated with the nutmeg oleoresin (Narasimhan and Dhake, 2006) present in the formulation of the active films. The values obtained comply with that established by the technical norms for the microbiological analysis of meat in Colombia.

CONCLUSIONS

The bovine loins covered with AFNO had a decreased pH, humidity percentage, peroxide values (POVs) and TVB-N values, conserving the red tones of the meat as storage time increased and

complied with the microbiological requirements established by the Colombian Technical Norms. Hence, the AFNO could be used to prolong the shelf life of bovine loins during the stages of storage and commercialization, enhancing the Colombian meat chain.

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Conflict of interest: The authors declare that there is no conflict of interest.

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